

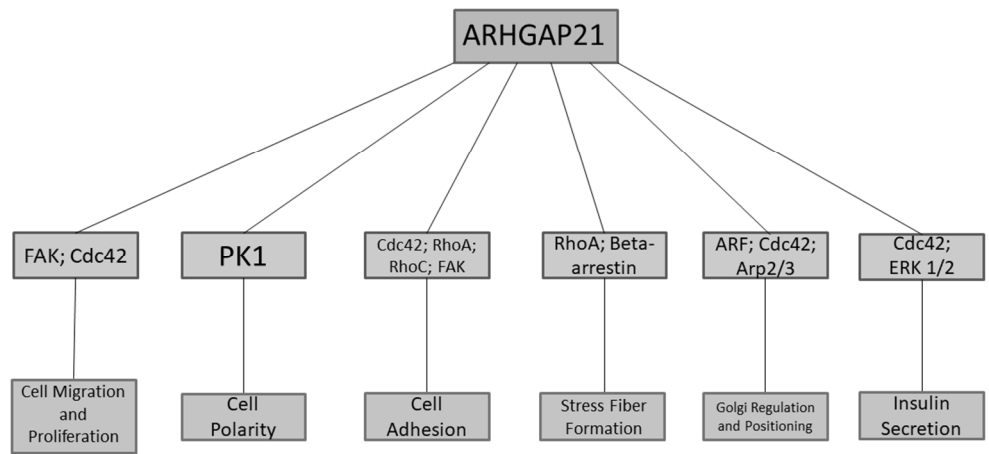


ARHGAP21 as a master regulator of multiple cellular processes

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ARHGAP21 and multiple roles it is implied in and the interactors through which it acts.

ARHGAP21 as a master regulator of multiple cellular processes

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Abstract

The cellular cytoskeleton is involved with multiple biological processes and it is tightly regulated by multiple proteins and effectors. Among these, the RhoGTPases family is one of the most important players, which are, in turn, regulated by many other elements. On the past decade, one of those regulators has been overlooked, despite being implied to have an important on many of those processes: ARHGAP21. In this paper, we aimed to review the literature available regarding ARHGAP21, in order to highlight its importance and the mechanisms of action that have been found so far for this still unknown protein involved with cell adhesion, migration, golgi regulation, cell trafficking and even insulin secretion.

Introduction

Ras is a superfamily of proteins in which the RhoGTPases family has a central role due to its importance in cytoskeleton-related processes as demonstrated by their interactions with Rho, Rac and Cdc42 (Hall, 2012). They act like molecular switches, cycling between a GDP-bound (Inactive) form and a GTP-bound (active) form. When GTP-bound, Rho GTPases interact with effectors, triggering cellular processes. The first and most important result of Rho GTPase signaling is the reorganization of the cellular cytoskeleton, mostly through their interaction with mDia and ROCK (Rho-associated kinase), as well as activation of Arp2/3 (Hall, 2012). The cytoskeleton, in turn, is an important element in multiple cellular processes, including migration, proliferation, adhesion, endocytosis and exocytosis.

Due to their importance, Rho GTPase activity is strictly regulated through three factors: guanine nucleotide exchange factors (Rho-GEFs), guanosine nucleotide dissociation inhibitors (Rho-GDIs) and Rho-GTPase activating proteins (Rho-GAPs) (Hodge and Ridley, 2016). GEFs lead to the exchange of GDP to GTP, acting as positive controllers that stimulate Rho GTPase action. GDIs act as negative controllers by sequestering the GDP-bound form of Rho GTPases, impairing GEFs's ability to activate them. Finally, GAPs also act as inhibitors of Rho-GTPase by promoting the catalysis of GTP into GDP, inactivating the Rho-GTPase after their activation by GEFs and promoting the cycling for correct Rho-GTPase function.

Among the multiple existing Rho-GAPs, this review will focus on ARHGAP21, an overlooked and understudied member of the family with documented relevancy in multiple biological processes ranging from cellular migration to insulin secretion. ARHGAP21 (also known as ARHGAP10) was identified by Bassères D.S. et al in 2002 (Basseres et al., 2002) and has a Rho-GAP domain, a pleckstrin homology (PH) domain, and a PDZ domain. Ever since, it has been the subject of only a few studies, despite being associated with multiple cellular processes (Dubois et al., 2005; Ferreira et al., 2015; Lazarini et al., 2013; Menetrey et al., 2007; Wang et al., 2012).

ARHGAP as a regulator of cytoskeletal processes in cancer cells

ARHGAP21's main interactors are Cdc42 (Dubois et al., 2005), RhoA (Lazarini et al., 2013) and ARFs, especially ARF1 and ARF6 (Klein et al., 2006; Menetrey et al., 2007) all of which are involved with cytoskeleton rearrangement and/or intracellular transport. These are the main proteins which ARHGAP21 interacts with and modulates in the multiple processes it is implied.

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3 A significant amount of the literature available refers to the role of ARHGAP21 in many cancer-
4 related subjects like migration and adhesion, indicating that the protein might be a tumoral
5 suppressor. ARHGAP21 has been identified as a cancer-related gene (Katoh and Katoh, 2004)
6 and is over-expressed in cancer cells (Carles et al., 2006). ARHGAP was found to interact with
7 Focal-Adhesion Kinase (FAK), with its depletion increasing cell migration (Bigarella et al., 2009).
8 The loss of ARHGAP21-FAK interaction led to increased FAK and Cdc42 activation and, thus, the
9 increase in migration (Bigarella et al., 2009). Thus, ARHGAP21 was identified as a potential
10 tumor suppressor, being able to control the progression of the disease. ARHGAP21 depletion
11 also increased cell migration, as well as decreased proliferation, in PC3 adenocarcinoma cell
12 lines through its activity as a Rho-GAP (Lazarini et al., 2013).
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16 A recently discovered signaling pathway, named “Lateral Signaling”, was also found to be
17 related with migration, with ARHGAP21 playing a key role (Zhang et al., 2016; Zhang and
18 Wrana, 2016). This lateral signaling pathway controls cell polarity and morphology which, in
19 turn, affect migration. This pathway consists of Prickle 1 (Pk1), an important cell polarity
20 protein, and ARHGAP21. ARHGAP21 interacts with Pk1, regulating RhoA (Zhang and Wrana,
21 2016). This mechanism is important to control the volatility of cell shape, which in turn for
22 regulates cell motility (Zhang and Wrana, 2016).
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26 Another important process on the development of cancer is cell adhesion – or, rather, the lack
27 of (Okegawa et al., 2004), and ARHGAP21 is also important for normal cell-cell adhesion.
28 ARHGAP21 moves from the nucleus to the cellular junctions during cell adhesion, reducing
29 Cdc42 activity (Barcellos et al., 2013). Knockdown of ARHGAP21 reduced the strength of cell-
30 cell adhesion and, in turn, increased migration (Barcellos et al., 2013). ARHGAP21 is a
31 necessary protein for α -tubulin acetylation, possibly by aiding the organization and
32 stabilization of tubulin, and pointed the lack of α -tubulin acetylation as a contributing factor
33 for the alterations of migration behavior in cells lacking ARHGAP21 (Barcellos et al., 2013). The
34 study, however, did not find an expected increase in epithelial-mesenchymal transition (EMT)
35 in ARHGAP21-lacking cells; instead, the ARHGAP21-less cells displayed reduced EMT. This is
36 due to a possible role of α -tubulin acetylation in EMT. ARHGAP21 is, therefore, important for
37 both adhesion and migration, and has an interesting relationship with α -tubulin (Barcellos et
38 al., 2013). This is supported by other studies, in which LYL1 knockdown mice led to a reduction
39 of ARHGAP21 and, consequentially, increased vascular permeability in the lungs of mice (Pirot
40 et al., 2014). Therefore, ARHGAP21 is implicated in multiple cytoskeleton-related processes,
41 like adhesion, proliferation and migration, through its interaction with Cdc42, RhoA, RhoC, and
42 FAK.
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47 Finally, ARHGAP21 is also involved in the formation of stress fibers (Anthony et al., 2011; Pirot
48 et al., 2014). In stress fiber formation by RhoA-mediated angiotensin stimulation, ARHGAP21
49 modulation of RhoA can be controlled by its interaction with beta-arrestin (Anthony et al.,
50 2011). When the beta-Arrestin/ARHGAP21 complex was disrupted, ARHGAP21 activity
51 increased, leading to RhoA inhibition and less stress fiber formation and actin reorganization
52 (Anthony et al., 2011).
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54 **ARHGAP21, cellular transport and Golgi**

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3 ARHGAP21 is also involved with the regulation of the Golgi apparatus and with the transport of
4 substances – endogenous or exogenous – within the cell. ARHGAP21 was identified as a link
5 between ARF and the downstream activation of Cdc42 in the regulation of Golgi function,
6 which then leads to the modulation of Arp2/3 and F-actin (Dubois et al., 2005). It has been
7 suspected for a long time that ARF coordinated Cdc42 and RhoA for the regulation of Golgi,
8 but the connection between them had eluded researchers until this work came out (D'Souza-
9 Schorey and Chavrier, 2006; Stamnes, 2002). The team was capable to demonstrate that
10 ARHGAP21 interacts with both ARF and Cdc42, establishing the link. A subsequent study
11 pointed that ARHGAP21 physically interacts with ARF through its PH domain and an adjacent
12 alfa-helix (Menetrey et al., 2007).
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15 ARHGAP21 depletion reduced Golgi's ability to be positioned correctly (Hehnly et al., 2010).
16 This positioning process is dependent on dynein and on microtubule, and is controlled by
17 Cdc42 which is, in turn, modulated by ARHGAP21, ARF1, and a vesicle-coating protein
18 coatomer.
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21 On top of its direct action over Golgi-related processes, ARHGAP21 is also involved in the
22 traffic of substances within the cell. The retrograde transport of *E. Coli's* Shiga toxin through
23 the secretory pathway is modulated by Rho GTPase activity, especially Cdc42 (Hehnly et al.,
24 2009). ARHGAP21 knockdown (or the constitutive activation of Cdc42) inhibited the transport
25 of Shiga toxin, and the addition of the toxin by itself lowered the levels of Cdc42 due to the
26 increase in ARHGAP21 – thus, ARHGAP21 seems to be important for the transport of the toxin
27 within the cell, through its modulation of Cdc42 (Hehnly et al., 2009). The transport of
28 Influenza Virus Neuraminidase was also modulated by ARHGAP21 (Wang et al., 2012). The
29 intracellular transport of influenza virus neuraminidase is crucial for the virus' life cycle, and
30 the depletion of ARHGAP21 (or the constitutive activation of Cdc42) stimulated neuraminidase
31 transport to the cell membrane. Silencing ARHGAP21 led to the increase of viral replication
32 (Wang et al., 2012). Thus, ARHGAP21 modulation over Cdc42 signaling appears to be
33 important in influenza virus infection through its influence over neuraminidase transport.
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38 **ARHGAP21 and insulin secretion**

39 Insulin secretion, specifically its second phase, in which secretion is less intensive but long-
40 lasting, is a process that depends on actin rearrangement (Kalwat and Thurmond, 2013). The
41 cortical actin acts as a barrier that prevents the passage of insulin granules coming from the
42 cytoplasm to the membrane and, thus, this cytoskeletal barrier needs to be rearranged.
43 Glucose stimulation of the pancreatic beta cell promotes that rearrangement through multiple
44 pathways, one of the most important involving Cdc42 (Nevins and Thurmond, 2003).
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47 Considering the involvement of the actin cytoskeleton in insulin secretion and the importance
48 of Rho GTPases in the modulation of actin (and the already established role of Cdc42 in insulin
49 secretion), it was not a stretch to imagine that ARHGAP21 could play a role in insulin secretion.
50 Thus, our group decided to investigate that possible involvement. We found is that ARHGAP21
51 is present in the beta cell (Ferreira et al., 2015), and ARHGAP21 knockdown increased basal
52 insulin secretion, but not glucose-stimulated insulin secretion, as expected. ARHGAP21
53 knockdown also led to an increase in ERK1/2 phosphorylation, which was previously implied in
54 actin rearrangement during insulin secretion (Kalwat and Thurmond, 2013; Longuet et al.,
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2005). These findings suggest, therefore, that ARHGAP21 seems to be involved in insulin secretion. However, its role as well as the underlying molecular mechanisms involved in this process remaining still unclear.

Concluding remarks

As we have shown in this review, ARHGAP21 appears to be an important protein in multiple processes, especially those linked with the actin cytoskeleton including migration, adhesion, intracellular transport and insulin secretion. Through ARHGAP21's modulation of Rho GTPases, especially RhoA and Cdc42, and its interaction with other proteins, like ERK and FAK, ARHGAP21 positions itself as a key member in the regulation of those processes. Despite its importance, ARHGAP21 has been a neglected protein since its discovery, and holds an incredible potential yet to be clarified by future studies.

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References

- Anthony DF, Sin YY, Vadrevu S, Advant N, Day JP, Byrne AM, Lynch MJ, Milligan G, Houslay MD, Baillie GS. 2011. beta-Arrestin 1 inhibits the GTPase-activating protein function of ARHGAP21, promoting activation of RhoA following angiotensin II type 1A receptor stimulation. *Molecular and cellular biology* 31(5):1066-1075.
- Barcellos KS, Bigarella CL, Wagner MV, Vieira KP, Lazarini M, Langford PR, Machado-Neto JA, Call SG, Staley DM, Chung JY, Hansen MD, Saad ST. 2013. ARHGAP21 protein, a new partner of alpha-tubulin involved in cell-cell adhesion formation and essential for epithelial-mesenchymal transition. *The Journal of biological chemistry* 288(4):2179-2189.
- Basseres DS, Tizzei EV, Duarte AA, Costa FF, Saad ST. 2002. ARHGAP10, a novel human gene coding for a potentially cytoskeletal Rho-GTPase activating protein. *Biochemical and biophysical research communications* 294(3):579-585.
- Bigarella CL, Borges L, Costa FF, Saad ST. 2009. ARHGAP21 modulates FAK activity and impairs glioblastoma cell migration. *Biochimica et biophysica acta* 1793(5):806-816.
- Carles A, Millon R, Cromer A, Ganguli G, Lemaire F, Young J, Wasyluk C, Muller D, Schultz I, Rabouel Y, Dembele D, Zhao C, Marchal P, Ducray C, Bracco L, Abecassis J, Poch O, Wasyluk B. 2006. Head and neck squamous cell carcinoma transcriptome analysis by comprehensive validated differential display. *Oncogene* 25(12):1821-1831.
- D'Souza-Schorey C, Chavrier P. 2006. ARF proteins: roles in membrane traffic and beyond. *Nature reviews Molecular cell biology* 7(5):347-358.
- Dubois T, Paleotti O, Mironov AA, Fraissier V, Stradal TE, De Matteis MA, Franco M, Chavrier P. 2005. Golgi-localized GAP for Cdc42 functions downstream of ARF1 to control Arp2/3 complex and F-actin dynamics. *Nature cell biology* 7(4):353-364.
- Ferreira SM, Santos GJ, Rezende LF, Goncalves LM, Santos-Silva JC, Bigarella CL, Carneiro EM, Saad ST, Boschero AC, Barbosa-Sampaio HC. 2015. ARHGAP21 prevents abnormal insulin release through actin rearrangement in pancreatic islets from neonatal mice. *Life sciences* 127:53-58.
- Hall A. 2012. *Rho family gtpases*. Portland Press Limited.

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2
3 Hehnlly H, Longhini KM, Chen JL, Stamnes M. 2009. Retrograde Shiga toxin trafficking is
4 regulated by ARHGAP21 and Cdc42. *Molecular biology of the cell* 20(20):4303-4312.
- 5 Hehnlly H, Xu W, Chen JL, Stamnes M. 2010. Cdc42 regulates microtubule-dependent Golgi
6 positioning. *Traffic (Copenhagen, Denmark)* 11(8):1067-1078.
- 7 Hodge RG, Ridley AJ. 2016. Regulating Rho GTPases and their regulators. *Nature reviews*
8 *Molecular cell biology* 17(8):496-510.
- 9 Kalwat MA, Thurmond DC. 2013. Signaling mechanisms of glucose-induced F-actin remodeling
10 in pancreatic islet beta cells. *Experimental & molecular medicine* 45:e37.
- 11 Katoh Y, Katoh M. 2004. Identification and characterization of ARHGAP27 gene in silico.
12 *International journal of molecular medicine* 14(5):943-947.
- 13 Klein S, Franco M, Chardin P, Luton F. 2006. Role of the Arf6 GDP/GTP cycle and Arf6 GTPase-
14 activating proteins in actin remodeling and intracellular transport. *The Journal of*
15 *biological chemistry* 281(18):12352-12361.
- 16 Lazarini M, Traina F, Machado-Neto JA, Barcellos KS, Moreira YB, Brandao MM, Verjovski-
17 Almeida S, Ridley AJ, Saad ST. 2013. ARHGAP21 is a RhoGAP for RhoA and RhoC with a
18 role in proliferation and migration of prostate adenocarcinoma cells. *Biochimica et*
19 *biophysica acta* 1832(2):365-374.
- 20 Longuet C, Broca C, Costes S, Hani EH, Bataille D, Dalle S. 2005. Extracellularly regulated
21 kinases 1/2 (p44/42 mitogen-activated protein kinases) phosphorylate synapsin I and
22 regulate insulin secretion in the MIN6 beta-cell line and islets of Langerhans.
23 *Endocrinology* 146(2):643-654.
- 24 Menetrey J, Perderiset M, Cicolari J, Dubois T, Elkhatib N, El Khadali F, Franco M, Chavier P,
25 Houdusse A. 2007. Structural basis for ARF1-mediated recruitment of ARHGAP21 to
26 Golgi membranes. *The EMBO journal* 26(7):1953-1962.
- 27 Nevins AK, Thurmond DC. 2003. Glucose regulates the cortical actin network through
28 modulation of Cdc42 cycling to stimulate insulin secretion. *American journal of*
29 *physiology Cell physiology* 285(3):C698-710.
- 30 Okegawa T, Pong RC, Li Y, Hsieh JT. 2004. The role of cell adhesion molecule in cancer
31 progression and its application in cancer therapy. *Acta biochimica Polonica* 51(2):445-
32 457.
- 33
34 Pirot N, Delpech H, Deleuze V, Dohet C, Courtade-Saidi M, Basset-Leobon C, Chalhoub E,
35 Mathieu D, Pinet V. 2014. Lung endothelial barrier disruption in Lyl1-deficient mice.
36 *American journal of physiology Lung cellular and molecular physiology* 306(8):L775-
37 785.
- 38 Stamnes M. 2002. Regulating the actin cytoskeleton during vesicular transport. *Current*
39 *opinion in cell biology* 14(4):428-433.
- 40 Wang S, Li H, Chen Y, Wei H, Gao GF, Liu H, Huang S, Chen JL. 2012. Transport of influenza virus
41 neuraminidase (NA) to host cell surface is regulated by ARHGAP21 and Cdc42 proteins.
42 *The Journal of biological chemistry* 287(13):9804-9816.
- 43 Zhang L, Luga V, Armitage SK, Musiol M, Won A, Yip CM, Plotnikov SV, Wrana JL. 2016. A lateral
44 signalling pathway coordinates shape volatility during cell migration. *Nature*
45 *communications* 7:11714.
- 46 Zhang L, Wrana JL. 2016. Regulation of Rho GTPases from the lateral sides of migrating cells.
47 *Small GTPases*:1-4.
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